

Skull identification key for Central European waterfowl (Aves: Anseriformes: Anatidae)

CHRISTIAN ELLROTT & GREGOR SCHMITZ

A b s t r a c t

Although the Anseriformes are among the most thoroughly studied bird orders in Central Europe, no precise key for the identification of their skulls was available. This paper seeks to remedy this lack by presenting a dichotomous identification key for 37 species of Anseriformes from Germany. Photographs and biometric measurements of the skulls are added to support the identification process.

K e y w o r d s : Aves, Anseriformes, Anatidae, skull morphology, identification key.

Z u s a m m e n f a s s u n g

Obwohl die Entenvögel (Anseriformes) zu den gut untersuchten Vogelgruppen zählen, lag bisher kein präziser Bestimmungsschlüssel für die Schädel der mitteleuropäischen Arten vor. Mit der vorliegenden Arbeit wird diese Lücke geschlossen und ein dichotomer Schlüssel zur Bestimmung von 37 in Deutschland lebenden Entenvogelarten vorgestellt. Zur Erleichterung der Bestimmung werden Fotos und Messdaten beigelegt.

C o n t e n t s

1	Introduction	347
2	Material and Methods	348
3	Identification key	352
4	References	362

1 Introduction

Although the Anseriformes are one of the most thoroughly studied bird orders, no identification key for the skulls of Central European species was yet available. As part of a graduate thesis project at Constance University (ELLROTT 2008), the skulls of 37 German waterfowl species were measured and analyzed with regard to functional anatomy, diet and feeding behavior. Based on the underlying data and measurements, a distinction can even be made between skulls of closely related species. In this paper we present a dichotomous identification key for all 37 species of the order. In addition, we include supportive morphometric data and illustrations.

The skull of the Anseriformes is desmognathous and holorhinous. It is further characterized by the following features: Processus angularis of the lower jaw strongly developed, joint area of the articulare with a distinct caudolateral to rostromedial ridge, rostropterygoid joint situated quite rostrally (features of Galloanseres); rostral end of palatina spatulate, rostropterygoid processus pedicellate, processus retroarticularis of the lower jaw sagittally compressed and knife-shaped, occipital fontanelles present, processus paracoroideus strongly developed (MICKOLEIT 2004).

GOODMANN & FISHER (1962) investigated the functional anatomy of the feeding apparatus of 17 species of the Anseriformes with seven species occurring in Central

Europe. In this work several measurements were presented ($n=1-28$).

BROWN et al. (2003) presented a skull identification key for selected species of birds, including Anseriformes. The key included general skull types and the bill-cranium-relation, and was supplemented by illustrations and some basic morphometric data. Unfortunately, the number of specimens examined was not specified (except when only one specimen was available). The illustrations were not very detailed and not comparable between species. Closely related species were shown with and without bill horn. Therefore, skulls that in reality are similar, appear to be very different. Finally, some important species are missing (e. g. *Mergellus albellus*) or represented in low numbers (e. g. *Branta bernicla* with $n=1$).

JANSEN & GESTEL (2009) present the following measurements on their “skullsite”: total length; length, width and height of cranium, upper bill length, skull relation (total length / upper bill length). The photographs are clear and informative. Unfortunately, the measurements are taken from just one specimen each. In addition, despite of the enormous number of included species, some Central European species are missing.

Because Germany has a good cross-section of the Central European fauna, we based our analysis on the species list in BARTHEL & HELBIG (2005), but we excluded very rare species in order to keep the identification key practicable (see chapter 2, “species selection”).

A c k n o w l e d g e m e n t s

We are grateful to Dr. DORIS MÖRIKE (Staatliches Museum für Naturkunde, Stuttgart), Dr. ELISABETH STEFAN (Landesdenkmalamt Baden-Württemberg, Arbeitsstelle Osteologie, Konstanz), BRITTA MÖLLENKAMP (Institut für Palaeoanatomie, Domestikationsforschung und Geschichte der Tiermedizin, Universität München) and Dr. S. FRAHNERT (Museum für Naturkunde, Berlin) for providing skull material. Dr. B. LEISLER (Max Planck Institut für Ornithologie, Radolfzell) helped us to define measurement methods and terminology. We also thank the referees for their comments and advice.

Tab. 1. Species included in the determination key. – Nomenclature according to BAUER et al. (2005), systematic order according to HOYO et al. (1992).

Scientific name	English name	German name
ANSERANATINAE		
Anserini		
<i>Cygnus olor</i> (J.K. Gmelin, 1789)	Mute Swan	Höckerschwan
<i>Cygnus atratus</i> (Latham, 1790)	Black Swan	Schwarzschwan
<i>Cygnus cygnus</i> (Linnaeus, 1758)	Whooper Swan	Singschwan
<i>Cygnus bewickii</i> Yarrell, 1830	Tundra Swan	Zwergschwan
<i>Anser fabalis</i> (Latham, 1787)	Bean Goose	Saatgans
<i>Anser brachyrhynchus</i> Baillon, 1834	Pink-footed Goose	Kurzschnabelgans
<i>Anser albifrons</i> (Scopoli, 1769)	Greater White-fronted Goose	Blässgans
<i>Anser erythropus</i> (Linnaeus, 1758)	Lesser White-fronted Goose	Zwerggans
<i>Anser anser</i> (Linnaeus, 1758)	Greylag Goose	Graugans
<i>Branta canadensis</i> (Linnaeus, 1758)	Canada Goose	Kanadagans
<i>Branta leucopsis</i> (Bechstein, 1803)	Barnacle Goose	Weißwangengans
<i>Branta bernicla</i> (Linnaeus, 1758)	Brent Goose	Ringelgans
<i>Branta ruficollis</i> (Pallas, 1769)	Red-breasted Goose	Rothalsgans
ANATINAE		
Tadornini		
<i>Alopochen aegyptiaca</i> (Linnaeus, 1766)	Egyptian Goose	Nilgans
<i>Tadorna ferruginea</i> (Pallas, 1764)	Ruddy Shelduck	Rostgans
<i>Tadorna tadorna</i> (Linnaeus, 1758)	Common Shelduck	Brandgans
Cairinini		
<i>Aix galericulata</i> (Linnaeus, 1758)	Mandarin Duck	Mandarinente
Anatini		
<i>Anas penelope</i> Linnaeus, 1758	Eurasian Wigeon	Pfeifente
<i>Anas stepera</i> Linnaeus, 1758	Gadwall	Schnatterente
<i>Anas crecca</i> Linnaeus, 1758	Common Teal	Krickente
<i>Anas platyrhynchos</i> Linnaeus, 1758	Mallard	Stockente
<i>Anas acuta</i> Linnaeus, 1758	Northern Pintail	Spießente
<i>Anas querquedula</i> Linnaeus, 1758	Garganey	Knäkente
<i>Anas clypeata</i> Linnaeus, 1758	Northern Shoveler	Löffelente
Aythyni		
<i>Netta rufina</i> (Pallas, 1773)	Red-crested Pochard	Kolbenente
<i>Aythya ferina</i> (Linnaeus, 1758)	Common Pochard	Tafelente
<i>Aythya nyroca</i> (Güldenstädt, 1770)	Ferruginous Duck	Moorente
<i>Aythya fuligula</i> (Linnaeus, 1758)	Tufted Duck	Reiherente
<i>Aythya marila</i> (Linnaeus, 1758)	Greater Scaup	Bergente
Mergini		
<i>Somateria mollissima</i> (Linnaeus, 1758)	Common Eider	Eiderente
<i>Clangula hyemalis</i> (Linnaeus, 1758)	Long-tailed Duck	Eisente
<i>Bucephala clangula</i> (Linnaeus, 1758)	Common Goldeneye	Schellente
<i>Melanitta fusca</i> (Linnaeus, 1758)	Velvet Scoter	Samtente
<i>Melanitta nigra</i> (Linnaeus, 1758)	Common Scoter	Trauerente
<i>Mergellus albellus</i> (Linnaeus, 1758)	Smew	Zwergsäger
<i>Mergus serrator</i> Linnaeus, 1758	Red-breasted Merganser	Mittelsäger
<i>Mergus merganser</i> Linnaeus, 1758	Goosander	Gänssäger
OXYURINAE		
Oxyurini		
<i>Oxyura jamaicensis</i> (J.K. Gmelin, 1789)	Ruddy Duck	Schwarzkopf-Ruderente

2 Materials und Methods

Species selection

Species selection is based on BARTHÉL & HELBIG (2005). From the 66 Anseriformes species listed in this paper, we excluded 29 species which (1) were previously recorded as introduced, arrived by assisted transport or escaped from captivity, and whose breeding population (if any) is thought not to be self-sustaining, or (2) were classified as vagrants with an average of less than five records per year since 1980.

Materials

The skull material originates mainly from the NIKOLAUS Collection (Staatliches Museum für Naturkunde, Stuttgart). We also used specimens from the following collections: Landesdenkmalamt Baden-Württemberg, Arbeitsstelle Osteologie, Konstanz; Institut für Palaeoanatomie, Domestikationsforschung und Geschichte der Tiermedizin, Universität München; Museum für Naturkunde, Berlin; SCHMITZ Collection (Universität Konstanz).

Of the 37 species selected, we measured ten specimens each, all of which met the necessary condition, i. e. no damage that could prevent proper measuring. See Tab. 1 for the nomenclature of the included species.

For photographic documentation we chose those individuals that were the most representative (i. e. with measurements close to mean values).

Abbreviations and measurements

The skulls were measured as follows (see also Fig. 1 and Tab. 2):

CL	Cranial length (measured from the middle of the fissura crano-facialis to the caudal end of the cranium)
Upper bill	
TOL	Tomium length
PL	Premaxillary length (measured between tip of upper bill and anterior end of nasus)
NL	Naris length
BWUB	Basal width of upper bill
UBS	Upper bill shape (= bill edge shape in dorsal view, divided into three categories: divergent (=D), parallel (=P) or convergent (=C) towards bill tip)
UBH	Upper bill height (= bill height in apical third)
Cranium	
CH	Cranial height
CW	Cranial width
SOW	Width of supraorbital isthmus
SG	Salt gland fossa (may be absent, weakly developed or distinct)
Lower bill	
LBL	Lower bill length
LBH	Lower bill height

In a few key couples we added (i. e. not included in Tab. 2) basal width of premaxillary (measured at the anterior end of nasus) and length (cranial – rostral) of lacrimal (Fig. 3).

Tab. 2. Measurements (in mm) of the treated 37 Anseriformes species ($n = 10$ per species; Max = maximum; Ø = Mean, Min = Minimum; SD = Standard deviation). – For abbreviations see chapter 2 and Fig. 1.

	Skull			Upper Bill						Cranium				Lower Bill		
	TL	UBL	CL	TOL	PL	NL	BWUB	UBS	UBH	CH	CW	SOW	SG	LBL	LBH	
<i>Cygnus olor</i>	Max	184.2	97.5	88.4	98.4	56.8	35.5	34.7		12.6	34.3	45.7	21.7		171.0	18.9
	Ø	178.6	93.3	85.3	95.0	52.8	31.7	33.3	P=100 %	11.0	32.8	44.0	20.1	Absent	165.7	17.7
	Min	172.1	89.1	82.1	91.8	49.7	27.8	31.0		10.2	31.3	41.9	17.6		159.0	16.4
	SD	4.9	3.1	2.0	2.3	2.1	2.1	1.2		0.8	0.8	1.3	1.4		4.1	0.8
<i>Cygnus cygnus</i>	Max	195.1	97.1	98.3	102.6	49.9	39.3	32.8		12.2	39.0	48.0	20.1		189.0	19.4
	Ø	180.0	88.9	91.0	93.4	44.3	35.4	30.5	P=100 %	10.6	36.8	44.9	17.9	Absent	169.5	17.2
	Min	167.5	79.5	83.2	86.0	38.9	32.9	28.1		8.8	34.8	42.7	15.3		152.8	15.6
	SD	8.8	5.0	4.2	5.3	3.0	1.8	1.5		1.0	1.3	1.6	1.3		10.7	1.1
<i>Cygnus bewickii</i>	Max	169.2	81.4	87.9	86.5	38.9	34.6	30.5		12.6	36.3	44.7	17.9		160.0	17.1
	Ø	156.5	74.4	82.1	80.1	36.5	30.4	29.0	P=100 %	10.0	34.3	41.6	15.5	Absent	146.1	15.7
	Min	144.7	69.3	75.4	74.4	33.6	26.9	27.7		8.0	32.5	38.3	13.8		132.7	14.0
	SD	7.5	4.3	3.5	4.3	2.0	2.2	1.0		1.3	1.2	1.9	1.3		8.6	0.9
<i>Anser fabalis</i>	Max	125.0	68.2	59.0	60.6	33.0	23.6	26.4		7.5	34.4	35.7	15.4		111.2	17.3
	Ø	116.9	61.4	55.5	55.2	29.5	21.8	24.4	C=100 %	6.7	32.4	33.2	13.5	Absent	102.3	15.8
	Min	110.8	56.8	53.3	50.1	27.1	19.4	22.8		6.1	30.9	26.5	11.1		93.9	15.1
	SD	5.8	4.0	2.1	3.6	2.0	1.2	1.1		0.5	1.1	2.5	1.3		5.5	0.7
<i>Anser brachyrhynchus</i>	Max	108.6	53.3	55.5	46.5	23.9	20.1	24.4		7.6	32.0	34.9	15.5		91.8	13.6
	Ø	100.7	48.2	52.5	43.1	21.4	17.8	22.0	C=100 %	6.4	30.0	31.8	13.2	Absent	85.9	12.4
	Min	92.8	43.2	49.7	39.2	19.0	15.5	18.5		5.2	28.1	28.9	10.0		78.4	11.8
	SD	5.0	3.0	2.3	2.2	1.5	1.5	1.7		0.7	1.4	1.8	1.5		4.5	0.6

	Skull			Upper Bill					Cranium				Lower Bill			
	TL	UBL	CL	TOL	PL	NL	BWUB	UBS	UBH	CH	CW	SOW	SG	LBL	LBH	
<i>Anser albifrons</i>	Max	110.0	55.3	55.8	50.3	25.8	19.5	24.1		6.8	30.7	32.3	13.8			
	Ø	104.7	51.8	52.9	46.8	24.1	18.9	23.0	C=100 %	6.2	29.5	31.1	12.4	Absent	94.7	13.3
	Min	98.7	48.3	50.4	43.8	22.8	17.6	21.7		5.1	27.3	29.6	10.8		82.8	11.2
	SD	3.4	2.1	1.6	2.1	1.1	0.6	0.7		0.5	0.8	0.9	0.9		3.5	0.5
<i>Anser erythropus</i>	Max	94.1	44.3	49.8	37.7	19.2	16.9	21.2		6.4	26.8	30.1	12.5		76.4	10.9
	Ø	88.5	40.5	48.0	35.0	16.7	15.8	19.5	C=100 %	5.5	25.9	28.7	11.3	Absent	72.8	10.0
	Min	85.1	38.1	45.5	32.6	15.1	14.7	17.3		4.9	24.9	27.1	9.8		68.1	9.2
	SD	3.0	2.0	1.6	1.7	1.1	0.8	1.2		0.4	0.6	0.9	0.7		2.6	0.6
<i>Anser anser</i>	Max	133.9	72.1	61.8	62.9	33.4	26.9	31.6		8.6	35.0	37.6	18.0		113.7	17.6
	Ø	124.7	65.2	59.5	58.3	30.6	23.6	28.6	C=100 %	8.2	33.4	36.1	15.0	Absent	107.6	16.5
	Min	117.6	60.0	57.0	55.3	28.7	21.6	25.7		7.8	31.6	34.2	13.8		102.3	15.6
	SD	4.4	3.4	1.5	2.2	1.4	1.6	1.6		0.3	1.1	1.0	1.2		2.7	0.5
<i>Branta canadensis</i>	Max	136.9	68.5	68.4	62.0	33.8	29.1	27.8		7.7	35.5	38.8	15.9		119.3	15.3
	Ø	128.5	63.9	64.6	59.3	29.0	25.6	25.2	C=70 %	7.0	33.4	36.4	13.5	Distinct or weak	113.6	14.2
	Min	118.8	58.8	59.9	53.1	25.4	23.7	20.5	P=30 %	6.1	31.9	33.8	11.8		104.3	12.8
	SD	4.7	2.8	2.9	2.6	2.3	1.8	1.9		0.5	1.1	1.3	1.2		4.2	0.6
<i>Branta leucopsis</i>	Max	91.5	40.1	51.9	34.2	16.9	16.5	19.0		6.0	28.3	31.2	12.8		76.1	10.4
	Ø	86.9	37.1	49.8	31.7	15.6	15.2	17.5	C=100 %	5.5	26.6	28.9	10.8	Weak or absent	71.2	9.8
	Min	83.2	34.5	48.1	29.7	14.3	13.8	16.5		4.8	25.5	27.5	9.0		68.3	8.8
	SD	2.8	1.7	1.3	1.4	0.7	1.0	0.7		0.4	0.9	1.1	0.9		2.7	0.5
<i>Branta bernicla</i>	Max	98.8	46.8	53.6	41.6	18.1	21.0	19.6		6.4	25.7	29.4	9.8		85.6	10.4
	Ø	92.1	41.6	50.5	37.1	16.5	18.5	18.4	C=100 %	5.6	24.9	28.4	8.7	Distinct	77.8	9.2
	Min	87.4	39.3	48.1	34.6	15.8	17.5	17.2		5.2	23.9	27.3	7.4		72.2	8.4
	SD	3.3	2.1	1.8	1.8	0.8	1.0	0.8		0.3	0.6	0.8	0.8		4.1	0.6
<i>Branta ruficollis</i>	Max	79.9	33.8	46.4	28.3	13.6	14.5	18.3		5.8	25.4	26.6	11.3		68.0	9.4
	Ø	76.8	32.0	44.8	26.3	12.8	13.5	16.8	C=100 %	5.4	24.0	25.1	9.8	Weak	62.2	8.5
	Min	72.2	29.4	42.7	23.2	11.0	12.6	15.4		4.7	23.2	24.1	8.7		58.7	8.3
	SD	2.7	1.4	1.4	1.4	0.7	0.7	0.9		0.3	0.7	0.9	0.8		2.6	0.3
<i>Alopochen aegyptiaca</i>	Max	115.8	53.4	62.5	53.0	30.4	18.9	22.4		8.2	29.1	32.4	14.8		100.9	11.9
	Ø	111.6	51.3	60.3	49.7	28.7	17.5	21.2	P=90 %	7.4	28.1	31.5	12.8	Absent	96.3	10.9
	Min	107.8	48.6	58.1	47.5	26.8	14.9	19.7	C=10 %	6.8	27.3	29.7	11.1		92.6	10.1
	SD	2.5	1.4	1.6	1.6	0.9	1.2	0.7		0.4	0.6	0.8	1.1		2.9	0.6
<i>Tadorna ferruginea</i>	Max	106.9	47.9	59.7	53.5	30.4	14.1	19.9		8.4	24.7	28.6	12.4		98.9	10.2
	Ø	101.9	45.1	56.8	49.9	28.0	13.4	18.4	P=100 %	7.2	23.9	27.5	10.9	Absent	92.3	9.7
	Min	94.1	41.7	52.4	44.9	24.8	12.6	17.2		6.3	23.2	25.6	9.6		84.4	9.1
	SD	3.7	1.7	2.1	2.4	1.6	0.5	0.8		0.6	0.5	0.8	1.0		4.1	0.3
<i>Tadorna tadorna</i>	Max	105.4	49.2	56.2	58.2	33.0	14.3	18.3		6.3	23.6	28.4	7.7		102.8	12.1
	Ø	97.7	44.5	53.2	53.4	29.7	12.8	17.1	D=100 %	5.2	22.7	26.9	7.1	Weak	93.9	10.3
	Min	89.1	39.8	49.3	48.1	26.4	11.2	15.7		3.8	21.8	25.3	6.1		84.9	9.3
	SD	4.9	2.6	2.4	3.1	1.8	0.9	0.8		0.7	0.7	0.9	0.5		5.3	0.8
<i>Aix galericulata</i>	Max	81.7	34.6	47.1	35.3	20.3	10.6	14.9		4.6	23.1	24.0	9.4		67.1	7.1
	Ø	78.2	33.0	45.2	33.3	19.1	9.8	14.2	C=80 %	4.3	22.7	23.2	8.8	Absent	64.2	6.7
	Min	74.5	30.3	43.0	30.7	17.0	9.3	13.8	P=20 %	3.7	22.0	22.2	7.4		61.3	6.3
	SD	2.1	1.1	1.3	1.3	0.8	0.4	0.3		0.3	0.3	0.7	0.6		1.8	0.3
<i>Anas penelope</i>	Max	89.1	39.4	49.7	41.2	26.6	10.1	16.1		5.4	22.7	23.9	6.6		78.6	8.7
	Ø	84.4	36.4	48.0	38.3	24.4	9.3	15.2	P=100 %	4.9	21.8	23.4	6.0	Weak	74.2	8.3
	Min	81.0	34.3	46.5	35.6	22.0	8.3	13.9		4.6	21.1	22.6	5.5		70.6	7.9
	SD	2.3	1.3	1.2	1.5	1.2	0.5	0.7		0.3	0.4	0.5	0.4		2.3	0.3

	Skull			Upper Bill					Cranium				Lower Bill		
	TL	UBL	CL	TOL	PL	NL	BWUB	UBS	UBH	CH	CW	SOW	SG	LBL	LBH
<i>Anas stepera</i>	Max	100.5	47.8	52.6	51.5	34.4	12.0	16.3		6.0	22.0	23.7	8.1		
	Ø	96.3	45.3	51.0	48.1	31.7	11.4	15.6	P=100 %	5.4	21.3	23.1	7.3	Absent	87.4 8.6
	Min	92.3	42.7	48.4	45.2	30.6	10.2	14.6		4.9	20.8	22.3	6.8		81.8 8.0
	SD	2.4	1.5	1.1	1.7	1.1	0.6	0.6		0.4	0.4	0.5	0.4		3.1 0.4
<i>Anas crecca</i>	Max	81.9	39.0	43.9	42.9	28.5	9.8	13.0		5.6	20.9	21.2	7.2		74.5 7.4
	Ø	79.0	36.5	42.5	39.9	26.3	8.5	12.5	P=100 %	4.9	20.2	19.6	6.4	Absent	70.4 6.7
	Min	76.2	34.2	41.3	37.9	24.4	7.5	11.2		4.5	19.0	18.3	5.1		68.1 6.3
	SD	1.9	1.5	0.8	1.5	1.2	0.7	0.5		0.3	0.5	0.8	0.6		2.1 0.3
<i>Anas platyrhynchos</i>	Max	117.4	57.6	59.8	62.6	38.2	15.7	21.3		8.6	27.2	28.9	10.6		107.3 10.6
	Ø	109.5	53.1	56.5	58.4	36.1	13.7	19.3	P=60 %	7.8	26.3	26.6	9.8	Absent	99.7 9.9
	Min	103.5	48.3	54.3	53.1	32.8	11.9	18.3	D=40 %	7.1	25.4	25.2	8.3		92.4 8.6
	SD	4.4	2.9	1.7	3.2	2.0	1.0	0.9		0.4	0.5	1.1	0.6		4.6 0.6
<i>Anas acuta</i>	Max	111.3	54.5	56.7	61.2	39.0	13.5	17.4		8.2	25.0	26.9	10.0		103.1 10.3
	Ø	104.5	50.6	53.9	55.9	35.9	12.4	16.5	P=50 %	6.7	23.8	25.8	9.0	Absent	96.0 9.6
	Min	95.9	45.8	50.2	51.1	32.6	10.8	15.7	D=50 %	5.2	23.0	23.7	8.2		86.9 8.8
	SD	4.4	2.5	2.0	3.0	1.8	0.9	0.5		1.0	0.5	1.0	0.6		4.5 0.4
<i>Anas querquedula</i>	Max	83.5	40.5	43.5	44.2	29.3	10.0	14.6		6.7	20.3	21.0	6.9		74.9 7.4
	Ø	80.3	37.8	42.4	41.8	27.0	9.5	13.8	P=90 %	5.6	19.8	19.9	6.2	Absent	71.9 6.7
	Min	76.6	36.1	40.3	39.5	24.7	8.7	13.1	D=10 %	4.9	19.4	18.3	5.6		69.3 6.2
	SD	1.9	1.2	0.9	1.4	1.2	0.4	0.5		0.6	0.3	0.9	0.4		1.6 0.3
<i>Anas clypeata</i>	Max	121.5	66.6	54.9	71.8	49.2	15.2	16.1		10.0	22.9	22.8	7.5		112.6 8.7
	Ø	112.8	60.8	52.0	66.0	43.8	13.9	15.2	D=100 %	8.2	21.8	21.9	6.8	Absent	104.8 8.1
	Min	106.2	56.9	49.3	59.2	40.3	12.2	14.3		6.8	21.0	21.1	5.3		99.3 7.4
	SD	5.2	3.4	2.0	4.2	2.9	0.8	0.6		0.9	0.5	0.6	0.7		4.8 0.4
<i>Netta rufina</i>	Max	104.1	53.3	51.9	53.8	32.5	15.7	24.4		5.5	23.7	27.3	8.9		93.7 9.8
	Ø	100.1	50.4	49.8	51.4	30.6	14.1	22.6	C=80 %	4.1	22.7	26.3	8.0	Absent (or weak)	89.3 9.2
	Min	94.4	46.9	47.5	48.2	27.9	12.8	21.4	P=20 %	3.3	21.8	25.4	6.9		84.4 8.7
	SD	2.9	1.9	1.3	1.7	1.4	0.8	0.9		0.7	0.5	0.6	0.5		2.5 0.3
<i>Aythya ferina</i>	Max	104.4	51.5	53.4	54.9	32.6	14.2	19.4		7.7	25.5	27.5	9.1		96.1 11.1
	Ø	99.2	47.3	51.9	51.0	30.1	13.1	18.7	P=100 %	6.7	24.7	26.9	7.9	Absent (or weak)	90.6 10.5
	Min	95.5	44.6	50.3	47.2	28.1	12.2	16.9		5.2	24.1	26.2	6.9		86.9 9.7
	SD	2.5	1.8	1.0	1.9	1.2	0.6	0.7		0.7	0.4	0.4	0.7		2.6 0.5
<i>Aythya nyroca</i>	Max	89.0	43.0	46.0	46.0	27.5	11.9	18.2		7.1	23.1	24.7	8.6		79.3 8.6
	Ø	85.4	40.9	44.5	43.9	25.9	10.8	16.5	P=80 %	5.8	22.2	23.7	8.2	Absent	76.2 7.9
	Min	80.7	38.3	42.4	40.9	24.2	9.8	13.8	D=20 %	5.1	21.5	22.8	7.8		71.0 7.2
	SD	2.3	1.2	1.1	1.4	0.9	0.6	1.3		0.6	0.5	0.5	0.2		2.2 0.4
<i>Aythya fuligula</i>	Max	97.0	45.9	51.1	50.8	27.8	13.7	22.0		7.6	24.1	26.1	8.3		88.6 10.9
	Ø	87.7	40.3	47.4	44.9	25.6	11.6	19.3	D=70 %	6.8	22.3	24.7	7.6	Absent or weak	78.4 9.0
	Min	83.9	37.4	44.8	41.9	23.9	10.7	17.9	P=30 %	6.2	21.2	23.9	7.2		74.6 8.3
	SD	3.4	2.1	1.5	2.2	1.1	0.8	1.1		0.5	0.8	0.7	0.3		3.7 0.7
<i>Aythya marila</i>	Max	99.1	46.1	53.0	51.5	28.5	13.8	21.5		8.9	24.5	28.8	7.5		91.6 11.7
	Ø	95.5	44.0	51.5	49.3	26.9	13.0	20.0	D=100 %	7.7	23.4	26.8	7.0	Distinct or weak	87.0 10.3
	Min	89.6	41.4	48.2	45.6	25.2	12.3	18.3		5.9	22.1	25.1	6.3		80.9 9.0
	SD	2.7	1.4	1.4	1.7	1.0	0.5	1.2		1.0	0.7	1.0	0.4		3.1 0.7
<i>Somateria mollissima</i>	Max	130.6	58.7	73.2	61.9	26.9	25.7	26.0		10.1	28.8	34.9	8.5		120.5 17.2
	Ø	125.7	56.0	69.7	59.1	25.7	23.0	24.0	C=50 %	8.7	27.7	32.6	7.8	Distinct	114.2 15.5
	Min	120.5	53.5	66.8	56.5	24.4	21.5	22.6	P=50 %	7.3	26.8	30.5	6.4		107.9 14.6
	SD	3.0	1.8	2.0	1.6	0.9	1.1	1.2		0.9	0.5	1.3	0.7		3.3 0.7

	Skull			Upper Bill					Cranium				Lower Bill			
	TL	UBL	CL	TOL	PL	NL	BWUB	UBS	UBH	CH	CW	SOW	SG	LBL	LBH	
<i>Clangula hyemalis</i>	Max	83.1	32.0	51.4	39.4	16.0	14.1	17.4		8.3	21.4	27.0	6.0		75.5	8.0
	Ø	79.5	30.6	48.9	37.2	15.2	13.0	16.5	P=100 %	7.4	20.6	25.7	5.2	Distinct	72.2	7.4
	Min	75.2	28.4	46.4	34.8	14.3	12.0	15.4		6.6	19.8	24.4	4.7		68.4	6.9
	SD	2.7	1.1	1.8	1.5	0.6	0.5	0.6		0.6	0.5	0.7	0.4		2.4	0.3
<i>Bucephala clangula</i>	Max	92.9	38.2	55.9	44.2	15.7	15.7	20.9		9.8	27.7	28.9	17.1		81.5	10.1
	Ø	86.1	34.4	51.7	40.7	14.4	13.7	18.9	P=80 %	8.6	25.5	27.0	13.1	Absent	75.6	8.8
	Min	80.5	31.4	48.2	37.6	13.2	12.2	16.7	C=20 %	7.1	23.2	25.6	11.2		69.8	7.9
	SD	4.8	2.3	2.7	2.7	0.7	1.0	1.4		0.8	1.5	1.2	2.0		4.8	0.8
<i>Melanitta fusca</i>	Max	114.7	55.8	59.5	62.7	28.8	17.1	27.0		8.3	27.4	30.3	10.3		106.8	12.8
	Ø	108.5	51.8	56.6	58.4	26.6	16.3	24.2	P=80 %	7.4	26.1	29.0	8.9	Distinct	99.7	11.4
	Min	100.3	47.3	53.1	53.8	23.9	15.5	21.8	D=20 %	6.6	24.8	27.2	7.7		92.8	10.2
	SD	5.5	3.5	2.2	3.5	1.8	0.5	1.9		0.7	1.0	1.1	0.7		5.4	0.9
<i>Melanitta nigra</i>	Max	103.7	50.8	52.9	56.2	24.9	14.8	22.6		7.9	26.4	28.3	8.5		96.8	11.1
	Ø	97.3	45.8	51.4	51.3	22.5	13.5	20.5	P=90 %	5.6	24.9	26.8	7.1	Distinct	89.8	10.6
	Min	91.8	41.5	48.9	47.3	20.7	12.5	17.0	C=10 %	4.1	23.8	25.3	6.0		82.6	9.8
	SD	4.0	2.9	1.2	3.0	1.4	0.8	1.7		1.2	0.7	0.9	0.7		4.5	0.4
<i>Mergellus albellus</i>	Max	78.9	38.3	41.6	35.0	17.5	11.9	12.1		2.6	20.6	25.2	7.6		70.3	8.3
	Ø	75.2	35.9	39.3	32.5	16.6	10.6	11.6	C=100 %	2.3	19.9	23.7	6.5	Absent	66.1	7.6
	Min	70.0	32.4	37.5	29.4	15.0	9.3	9.8		2.0	19.4	22.7	5.8		59.2	5.8
	SD	2.9	1.8	1.2	1.7	0.8	0.9	0.7		0.2	0.4	0.7	0.6		3.5	0.7
<i>Mergus serrator</i>	Max	118.5	72.3	47.4	66.8	45.0	17.1	14.5		2.5	20.4	30.0	7.5		114.4	8.6
	Ø	108.4	64.0	44.4	60.2	39.1	14.7	13.0	C=100 %	2.3	19.9	27.3	6.6	Absent	103.5	7.7
	Min	97.7	56.0	41.7	51.4	34.1	13.2	11.5		2.1	19.2	25.5	6.0		90.4	6.9
	SD	6.7	5.2	1.7	4.9	3.4	1.1	0.9		0.2	0.4	1.2	0.5		7.3	0.5
<i>Mergus merganser</i>	Max	124.0	70.6	54.1	64.4	36.8	18.3	17.5		2.8	22.3	32.2	12.5		117.4	10.6
	Ø	115.6	64.4	51.1	59.5	33.6	16.4	15.3	C=100 %	2.3	21.1	30.7	11.4	Mostly weak	109.0	9.1
	Min	103.4	55.6	47.0	51.6	28.7	14.1	13.0		1.9	19.8	29.1	9.4		96.3	7.5
	SD	8.2	5.6	2.7	5.2	3.0	1.4	1.6		0.3	0.8	1.1	0.9		8.0	1.1
<i>Oxyura jamaicensis</i>	Max	85.7	40.7	45.7	43.9	22.4	15.8	18.5		7.5	22.5	23.5	8.5		77.1	9.7
	Ø	82.7	38.1	44.6	41.7	21.2	14.4	17.5	D=100 %	6.1	21.1	22.8	6.2	Absent	74.6	8.7
	Min	79.8	36.2	43.6	39.8	20.0	13.0	16.5		4.9	20.2	22.3	5.5		72.5	8.1
	SD	1.6	1.3	0.7	1.1	0.8	0.7	0.6		0.7	0.6	0.5	0.8		1.4	0.4

3 Identification key

The complete data set of all 37 species (each based on 10 specimens) is presented in Tab. 2.

- 1 Upper bill narrow, <9 mm wide (measured in the middle of the bill between tip and naris). (M ergini) 2
- Upper bill broadened, duck- or goose-like, >9 mm wide.... 4
- 2 Premaxillary anterior to naris relatively short: PL < 1.5 times NL (Fig. 5a); UBL < 40 mm. – Fig. 44. *Mergellus albellus*
- Premaxillary anterior to naris relatively long: PL > 2 times NL (Fig. 5b, c); UBL > 40 mm. 3
- 3 SOW < 7.5 mm; PL (Ø 39.1 mm) about 2.5 times NL (Ø 14.7 mm); bill tip slightly down curved (Fig. 5c). – Fig. 45. *Mergus serrator*
- SOW > 9 mm; PL (Ø 33.6 mm) about 2 times NL (Ø 16.4 mm); bill tip often distinctly down curved (Fig. 5b). – Fig. 46. *Mergus merganser*
- 4 UBL > 140 mm and length of lacrimal (cf. Fig. 3) > 25 mm. [In large geese (*Anser*), the cross-section of the premaxil-

lary is thickened and spongy, whereas in *Cygnus* it is flattened; *Cygnus atratus* (only a single skull studied): UBL ca. 142 mm, length of lacrimal ca. 23 mm, SG distinct]. (A ns er i n i : *Cygnus*) 5

- UBL < 140 mm and length of lacrimal < 23 mm. 7
- 5 Braincase flattened in lateral view, dorsal roof of the braincase with a slight transverse depression dorsally (Fig. 3a); protrusion above the bill usually present (particularly in males); processus orbitalis of lacrimal without rostral extension. – Fig. 8. *Cygnus olor*
- Braincase rounded in lateral view and without a transverse depression dorsally (Fig. 3b); protrusion above the bill absent or slightly developed; processus orbitalis of lacrimal with rostral extension. 6
- 6 Bill long: TOL mostly > 85 mm (86–102 mm), PL 38.9–50.0 mm; processus orbitalis of lacrimal with a wide constriction (width of constriction 6.5–10.0 mm) (Fig. 3b). – Fig. 9. *Cygnus cygnus*
- Bill shorter: TOL mostly < 85 mm (75–86 mm), PL 33.6–38.9 mm; processus orbitalis of lacrimal with a narrow

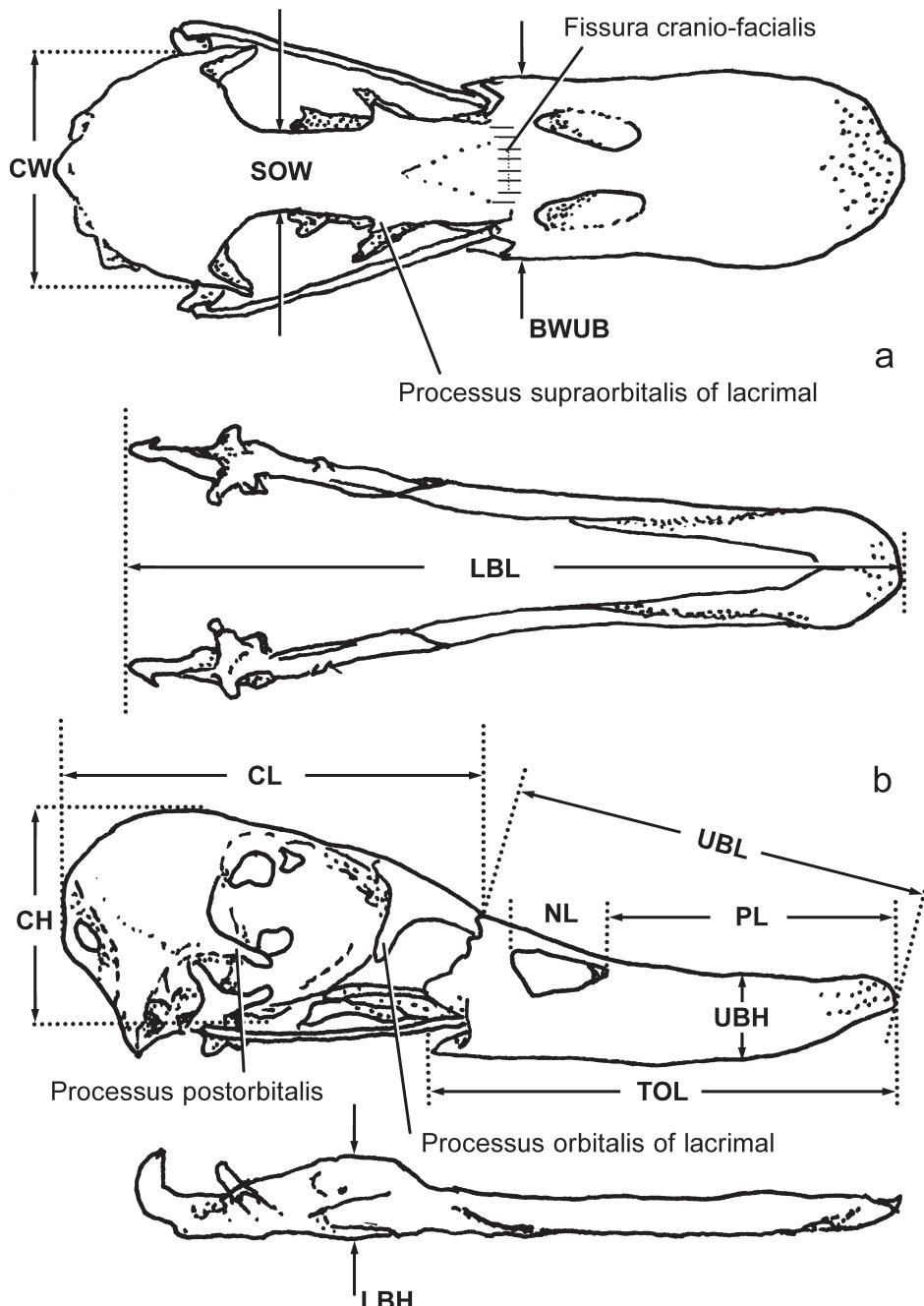
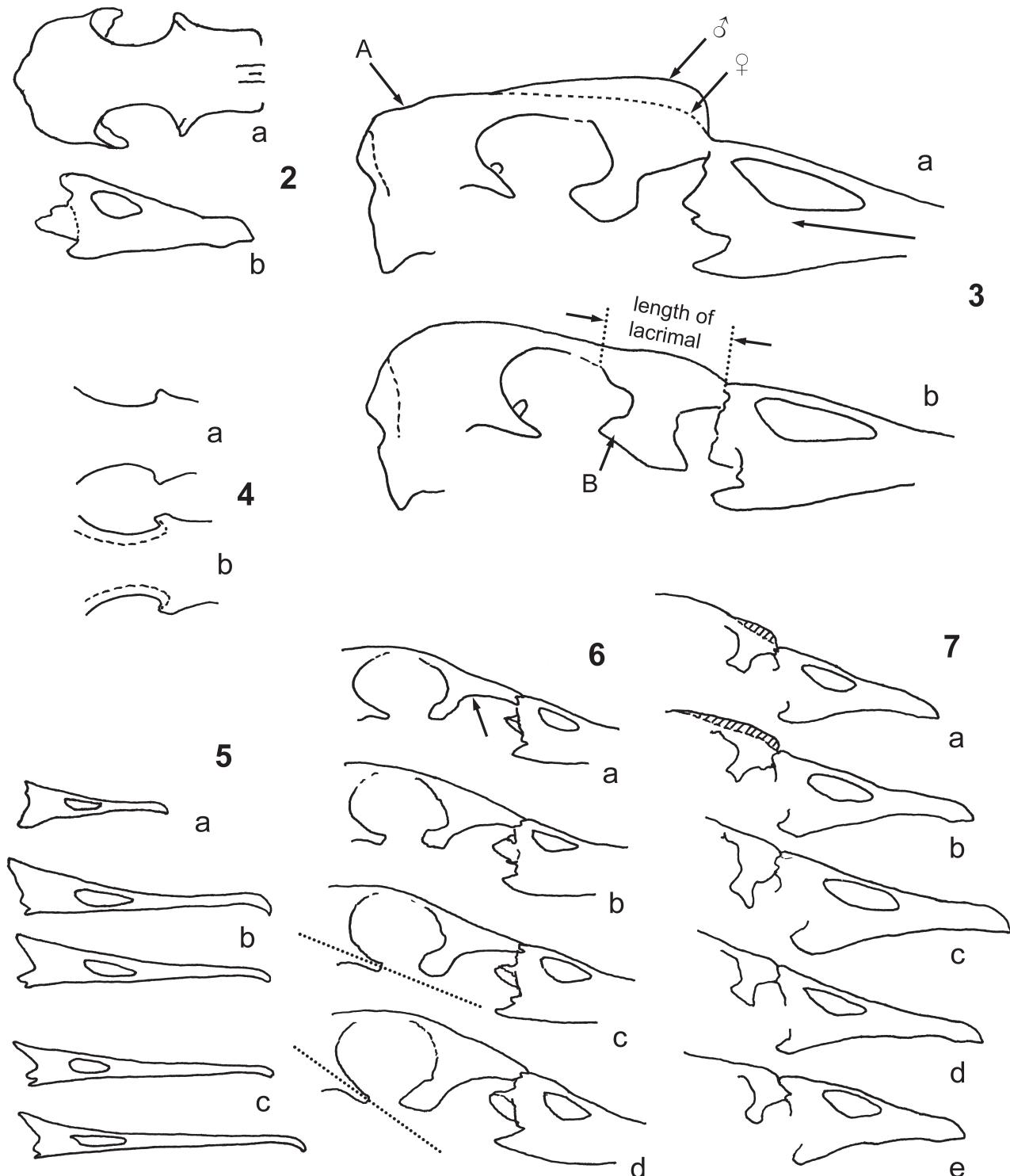


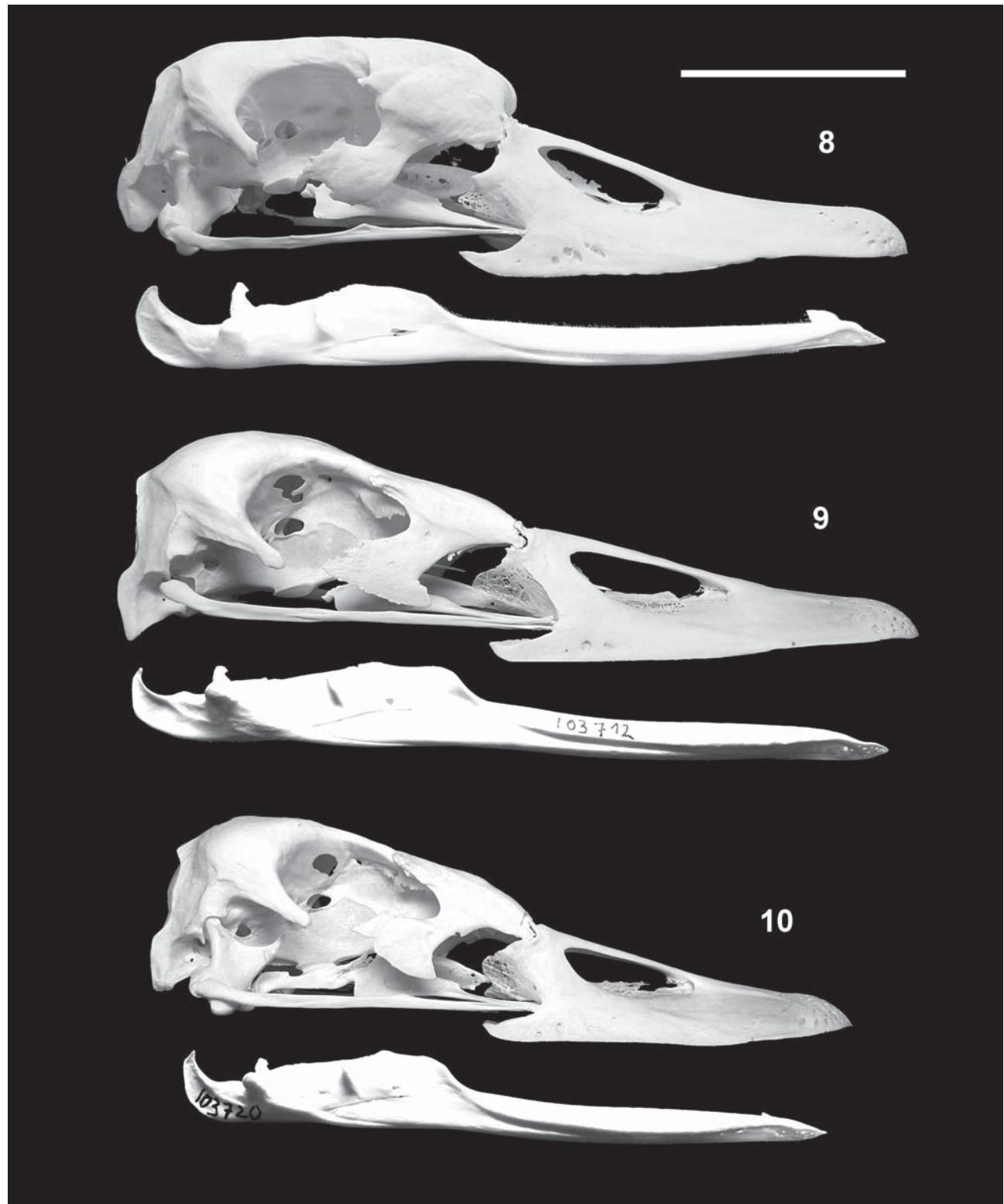
Fig. 1. Measurements of a duck skull (*Aythya fuligula*) in (a) dorsal and (b) lateral view. – For abbreviations see chapter 2.

- constriction (width of constriction 4.5–7.0 mm). – Fig. 10. *Cygnus bewickii*
- 7 Skull with at least two of the following three features: Edges of upper bill convergent in dorsal view; lower edge of upper bill in lateral view concave over nearly its entire length; premaxillary anterior to naris entirely filled with inner spongy bone structure and bill tip curved rostrally. (A n s e r i n i) 8
- Skull with at least two of the following three features: Edges of upper bill parallel or divergent in dorsal view; lower edge of upper bill straight or convex in lateral view; premaxillary anterior to naris single-layered at least in some lateral parts. 16
- 8 Processus supraorbitalis of lacrimal (cf. Fig. 1) slightly developed or blunt; SG always absent (Fig. 4a) (*Anser*). [Processus supraorbitalis may be distinct in domestic breeds of *Anser anser*, wild relatives have a maximum TL of 135 mm.] 9

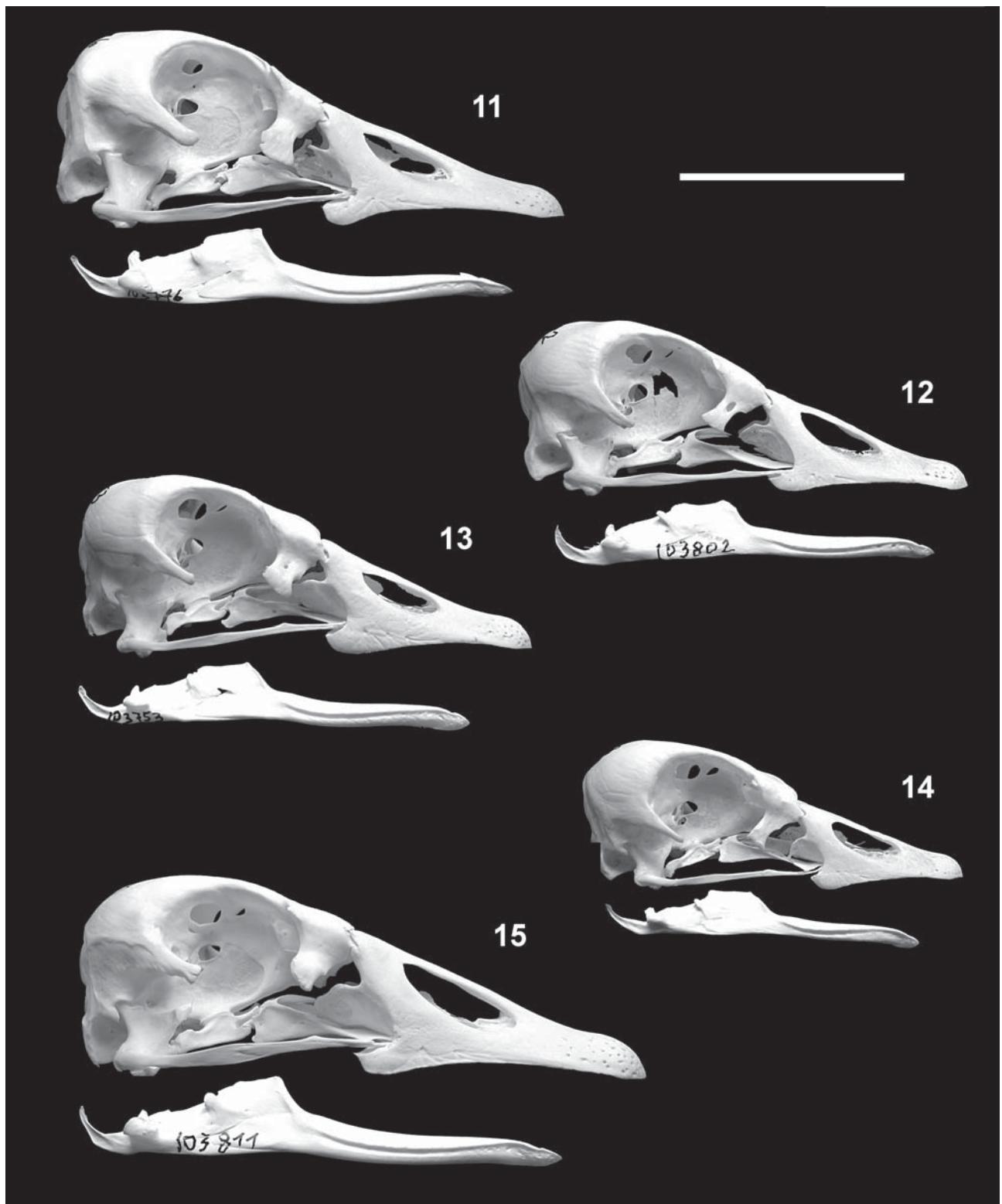
- Processus supraorbitalis of lacrimal distinct and curved backwards; SG distinct or slightly developed (Fig. 4b). (Branta) **13**
- 9** Frontal region posterior to the fissura crano-facialis medially slightly bulged in the midline (Fig. 7a, b). **10**
- Frontal region posterior to the fissura crano-facialis straight or concave (Fig. 7c–e). **11**
- 10** TOL 32–38 mm (see also Fig. 7a). – Fig. 14. *Anser erythropus*
- TOL 43–51 mm (see also Fig. 7b). – Fig. 13. *Anser albifrons*
- 11** PL 19–24 mm; culmen above the nares convex (Fig. 7e). – Fig. 12. *Anser brachyrhynchus*
- PL > 26 mm. **12**
- 12** Basal width of premaxillary 15–18 mm (measured at the anterior end of nares); NL 19–24 mm (Fig. 7d). – Fig. 11. *Anser fabalis*
- Basal width of premaxillary 18–21 mm; NL 21–27 mm (Fig. 7c). – Fig. 15. *Anser anser*
- 13** TOL > 50 mm. – Fig. 16. *Branta canadensis*
- TOL > 45 mm. **14**
- 14** TL < 80 mm. – Fig. 19. *Branta ruficollis*
- TL > 80 mm. **15**
- 15** SG distinct; CH 23–26 mm. – Fig. 18. *Branta bernicla*
- SG weakly developed or absent; CH 25–29 mm. – Fig. 17. *Branta leucopsis*
- 16** Processus supraorbitalis of lacrimal prominent, spine-shaped (Figs. 36, 43); edges of upper bill divergent and NL > 12 mm; SG mostly distinct. (Mergini without *Mergus*, *Mergellus*, and *Bucephala*) **17**
- Processus supraorbitalis of lacrimal not prominent (if prominent, as occasionally in *Aythya marila* and *A. fuligula*, then edges of upper bill divergent and NL < 15 mm); SG mostly absent. **19**
- 17** TL < 85 mm. – Fig. 37. *Clangula hyemalis*
- TL > 120 mm. – Fig. 36. *Somateria mollissima*
- TL 90–115 mm. (*Melanitta*) **18**
- 18** Minimum distance between nostrils > 2.5 mm; males with lateral protuberances near bill base. – Figs. 40, 41. *Melanitta fusca*
- Minimum distance between nostrils < 2.5 mm; males with a pair of distinct protuberances between the nares and the fissura crano-facialis. – Figs. 42, 43. *Melanitta nigra*
- 19** Processus orbitalis of lacrimal elongated, sword-shaped, reaches or extends beyond the vertical axis through the centre of the orbit; premaxillary anterior to nares nearly as long as nares. – Figs. 38, 39. *Bucephala clangula*
- Processus orbitalis of lacrimal thin or with broadened tip, never reaches the vertical axis through the centre of the orbit; premaxillary anterior to nares at least 1.5 times NL. ... **20**
- 20** Upper bill flattened, its edges divergent and bill tip conspicuously upturned; PL ca. 1.5 times NL; processus orbitalis of lacrimal short, blunt; posterior edge of orbit conspicuously rounded. – Fig. 47. *Oxyura jamaicensis*
- Upper bill shaped differently; PL at least 2 times NL; processus orbitalis of lacrimal elongated; posterior edge of orbit (except in *Tadorna tadorna*) not conspicuously rounded. **21**
- 21** Bill narrow, i. e. < 13 mm wide in rostral third of the bill and bill edges convergent; SOW < 7.3 mm. – Fig. 23. *Aix galericulata*
- Bill relatively wide, i. e. width in rostral third of the bill > 13 mm (if < 13 mm, then upper bill edges not convergent); SOW > 7.3 mm. **22**
- 22** Upper and lower bill entirely upturned; posterior edge of orbit conspicuously rounded. – Fig. 22. *Tadorna tadorna*
- Upper and lower bill not conspicuously upturned; posterior edge of orbit not conspicuously rounded. **23**
- 23** Braincase stout, CW > 29 mm; bill goose-like, i. e. edges slightly convergent and curved at bill tip; front very wide in lacrimal area (Fig. 2). – Fig. 20. *Alopochen aegyptiaca*
- Braincase less stout, CW < 29 mm; bill duck-like, i. e. ± flattened at bill tip (sometimes slightly curved in nail region); front less wide in lacrimal area. **24**
- 24** Processus orbitalis of lacrimal narrow, its edges parallel or pointed apically. (Aythini) **25**
- Processus orbitalis of lacrimal apically plane and broadened. (Anatin + *Tadorna ferruginea*) **29**
- 25** Upper bill very flat, edges convergent, bill tip acuminate, BWUB > 21 mm. – Fig. 31. *Netta rufina*
- Upper bill edges parallel or divergent, BWUB < 21 mm. (Aythya) **26**
- 26** Upper bill edges parallel and TOL > 47 mm. – Fig. 32. *Aythya ferina*
- Upper bill edges divergent (if parallel then TOL < 47 mm). ... **27**
- 27** Maximum upper bill width < 20 mm; ridge between nares and bill tip rather flattened, < 7.1 mm high. – Fig. 33. *Aythya nyroca*
- Maximum upper bill width > 20 mm; ridge between nares and bill tip less flattened, usually > 7.1 mm high. **28**
- 28** TL < 89 mm; width of fissura crano-facialis < 11 mm; SG absent or very slightly developed; processus supraorbitalis of lacrimal mostly small. – Fig. 34. *Aythya fuligula*
- TL > 89 mm; width of fissura crano-facialis > 11 mm; SG usually conspicuous; processus supraorbitalis of lacrimal mostly elongated. – Fig. 35. *Aythya marila*
- 29** TL < 90 mm. **30**
- TL > 90 mm. **32**
- 30** CW > 22 mm, CH > 21 mm; processus supraorbitalis of lacrimal distinct. – Fig. 24. *Anas penelope*
- CW < 22 mm, CH < 21 mm; processus supraorbitalis of lacrimal short. **31**
- 31** Upper bill slightly diverging; maximum upper bill width 13.6–16.3 mm. – Fig. 29. *Anas querquedula*
- Upper bill edges parallel; maximum upper bill width 12.3–13.6 mm. – Fig. 26. *Anas crecca*
- 32** Bill extremely widened rostrally; maximum bill width 27–33 mm. – Fig. 30. *Anas clypeata*
- Bill edges at most slightly divergent; maximum bill width < 25 mm. **33**
- 33** CH < 22.5 mm; rostral ventral edge of lacrimal and processus orbitalis meet at a distinct angle (ca. 100°, see arrow in Fig. 6a). – Fig. 25. *Anas stepera*
- CH > 22.5 mm; rostral ventral edge of lacrimal and processus orbitalis meet in a slight curve (Fig. 6b–e). **34**
- 34** Dorsal contour of lacrimal slightly convex in lateral view; longitudinal axis of processus postorbitalis directed below the posterior extension of the tomium (Fig. 6d); PL < 31 mm. – Fig. 21. *Tadorna ferruginea*
- Dorsal contour of lacrimal straight or protruding only posteriorly in lateral view; processus postorbitalis projects more caudally towards the posterior extension of the tomium (Fig. 6c); PL > 31 mm. **35**
- 35** BWUB > 18 mm; dorsal contour of lacrimal straight in lateral view (Fig. 6c). – Fig. 27. *Anas platyrhynchos*
- BWUB < 18 mm; dorsal contour of lacrimal slightly convex in lateral view (Fig. 6b). – Fig. 28. *Anas acuta*



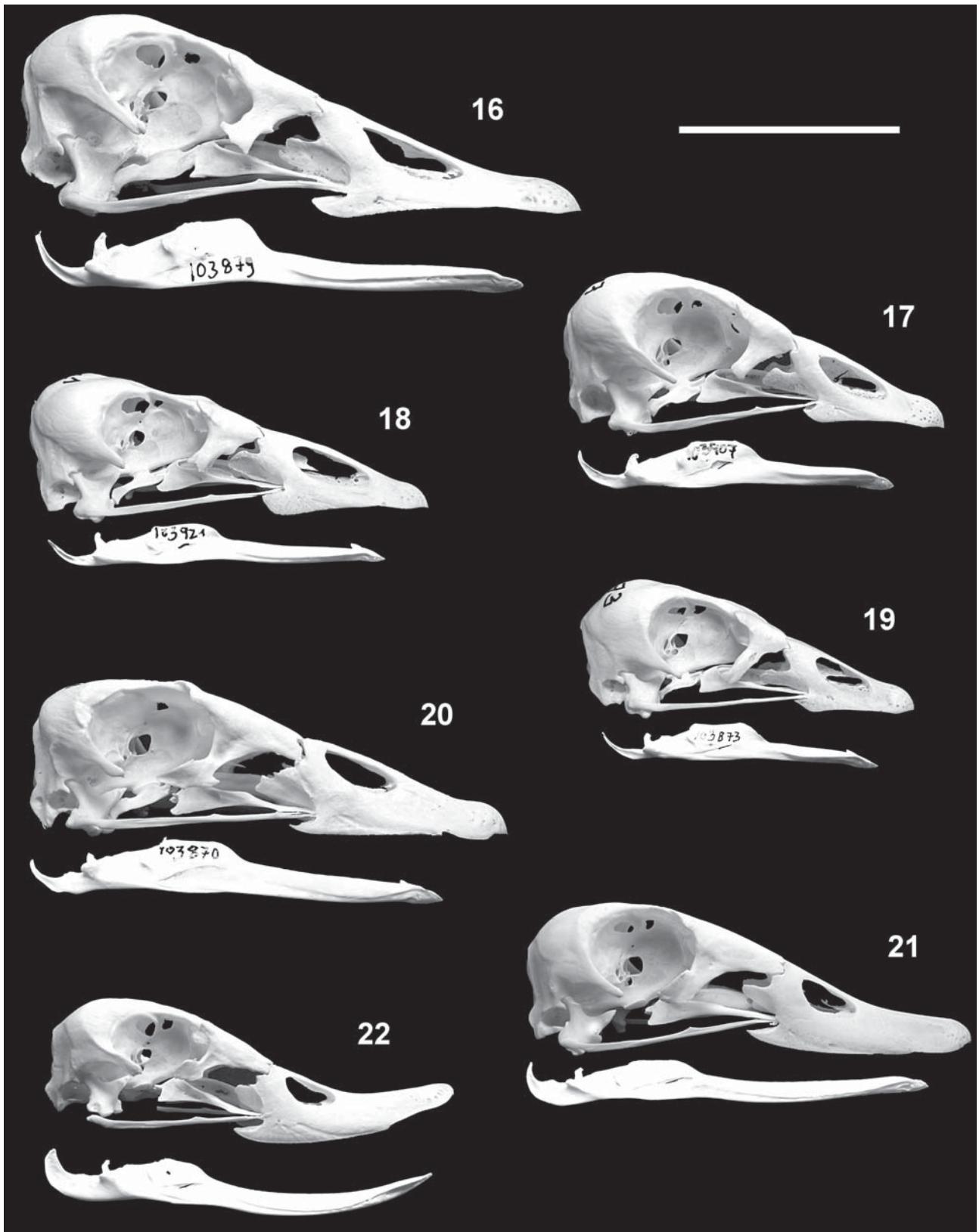
Figs. 2–7. Details of skulls of Anseriformes. – 2. *Alopochen aegyptiaca*, (a) cranium in dorsal view, (b) upper bill. 3. Cranium and basal part of upper bill of (a) *Cygnus olor* and (b) *C. cygnus* [A = transverse depression, B = rostral extension of processus orbitalis]. 4. Interorbital isthmus, (a) without (*Anser*) and (b) with (*Branta*) salt gland fossa. 5. Upper bills of (a) *Mergellus albellus*, (b) *Mergus merganser* (with variation), (c) *M. serrator* (with variation). 6. Lacrimal region of (a) *Anas stepera*, (b) *A. acuta*, (c) *A. platyrhynchos*, (d) *Tadorna ferruginea*. 7. Upper bill of (a) *Anser erythropus*, (b) *A. albifrons*, (c) *A. anser*, (d) *A. fabalis*, (e) *A. brachyrhynchus*.



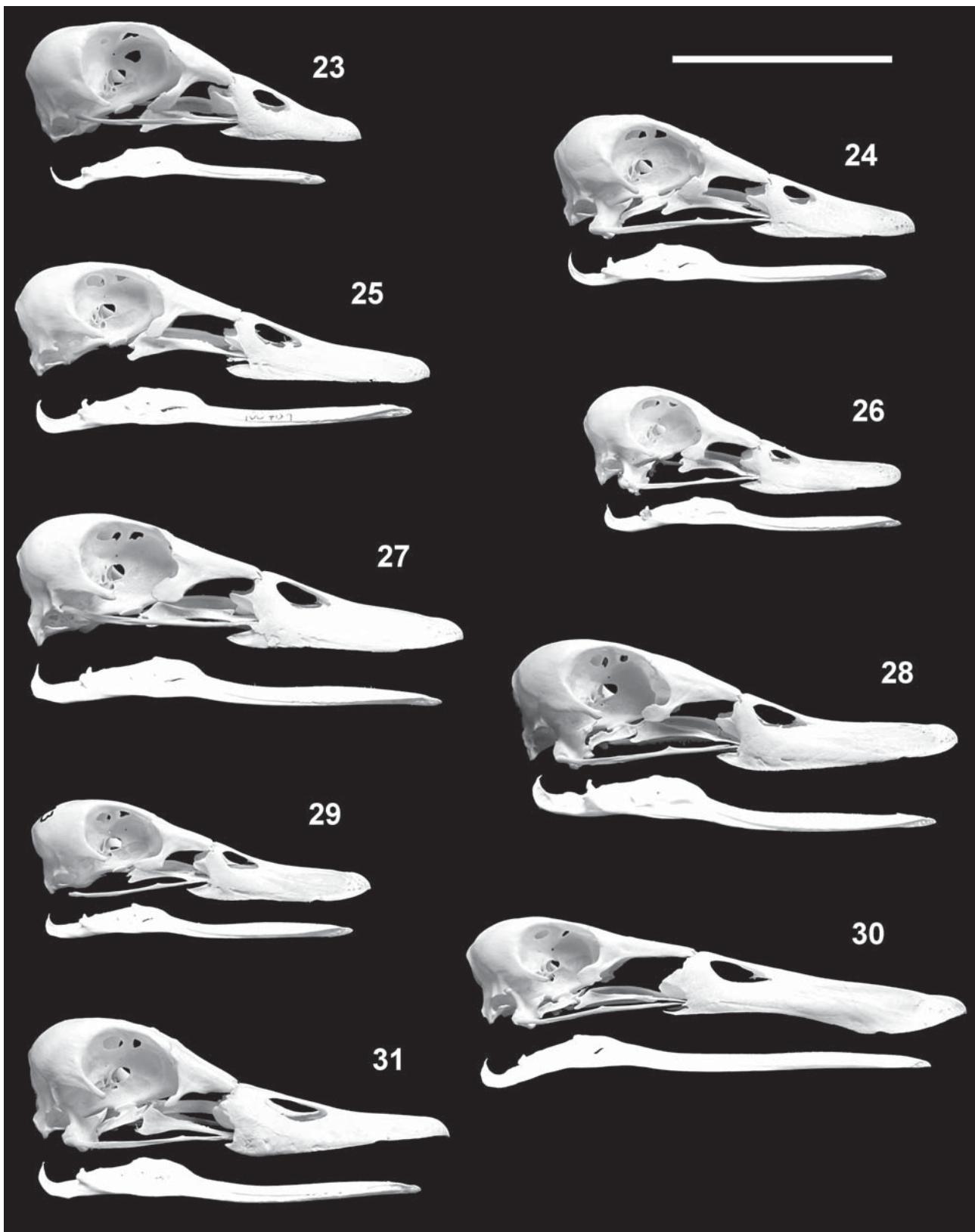
Figs. 8–10. Skulls of Anseriformes, lateral view. – 8. *Cygnus olor*. 9. *C. cygnus*. 10. *C. bewickii*. – Scale: 5 cm.



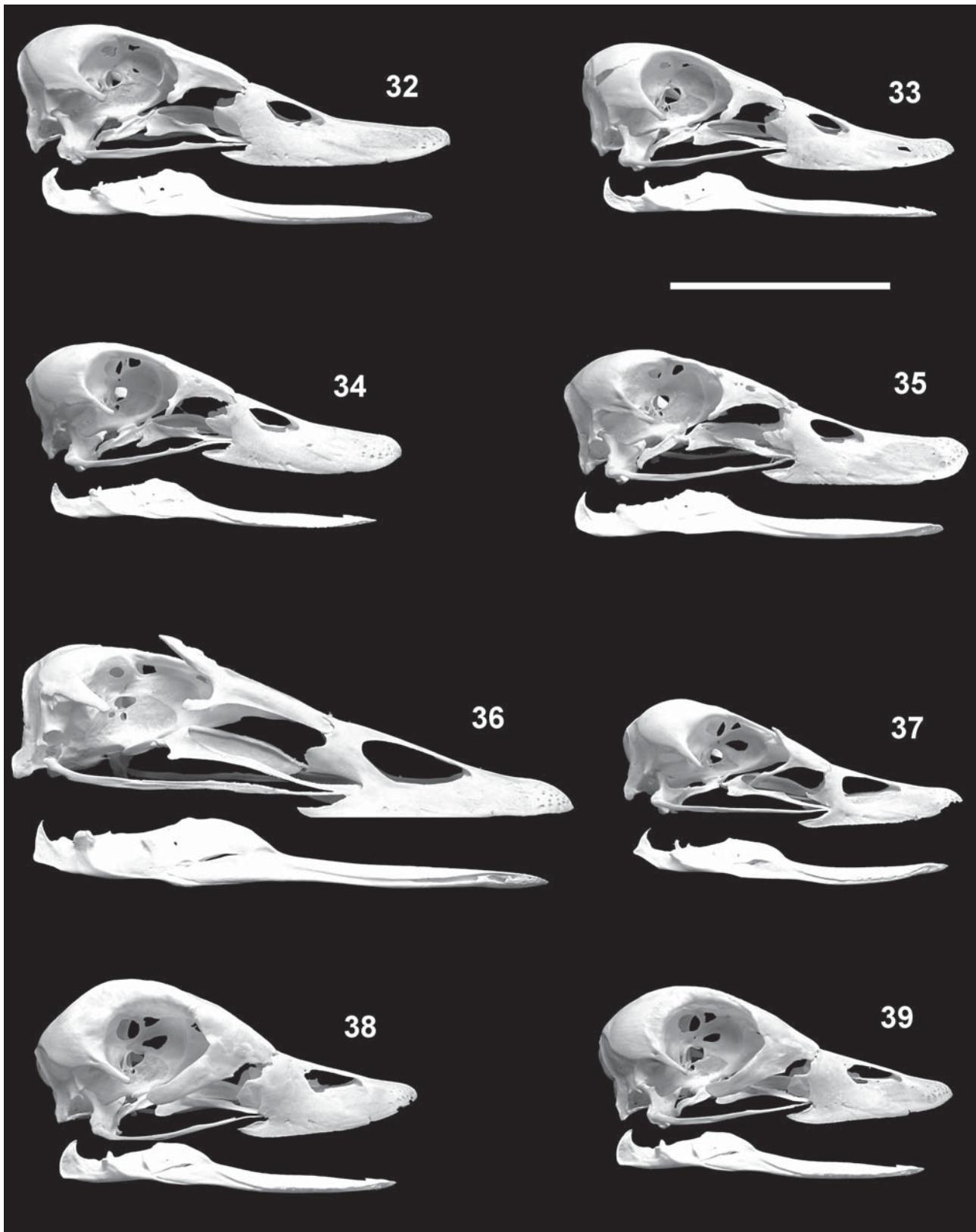
Figs. 11–15. Skulls of Anseriformes, lateral view. – 11. *Anser fabalis*. 12. *A. brachyrhynchus*. 13. *A. albifrons*. 14. *A. erythropus*. 15. *A. anser*. – Scale: 5 cm.



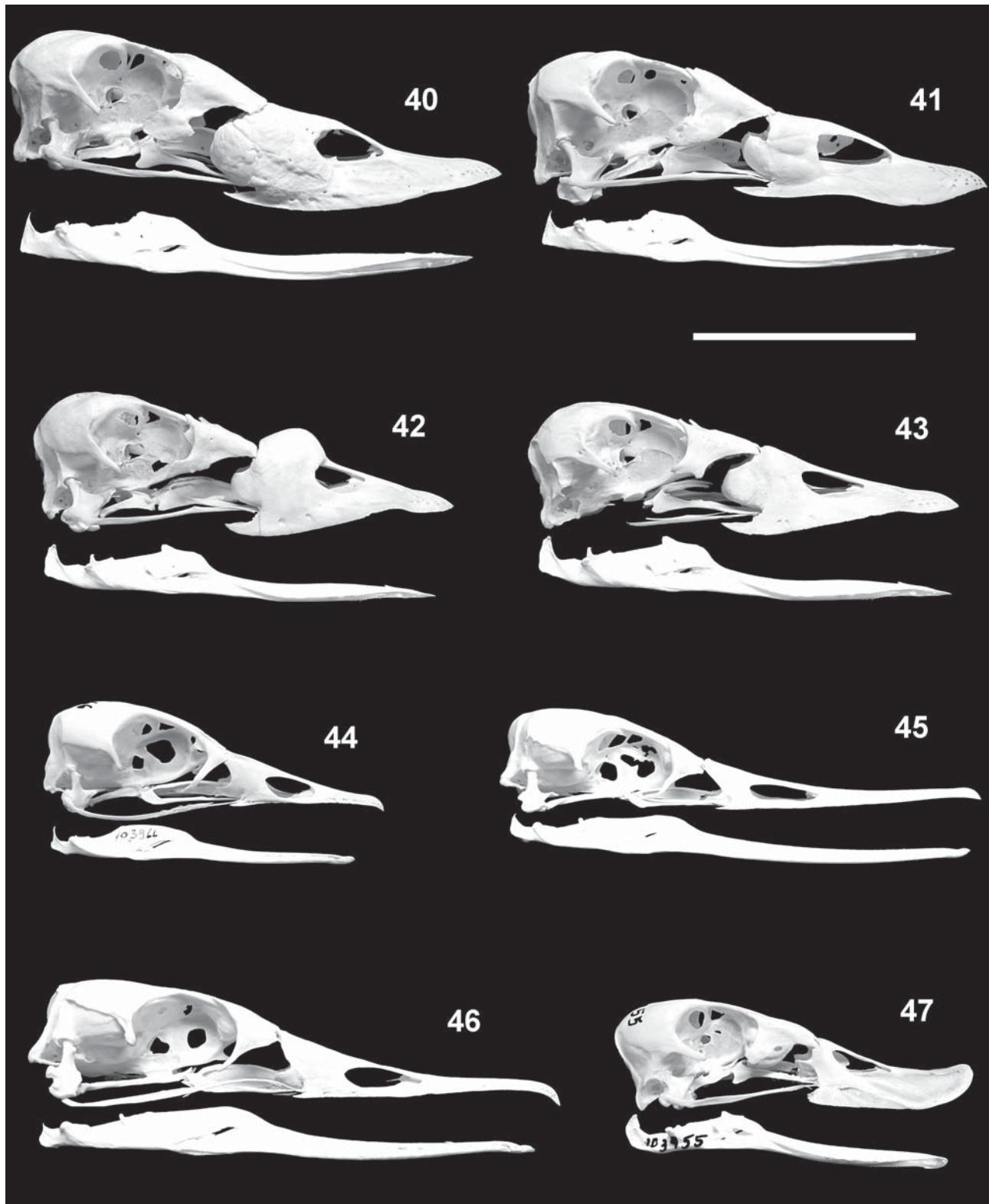
Figs. 16–22. Skulls of Anseriformes, lateral view. – **16.** *Branta canadensis*. **17.** *B. leucopsis*. **18.** *B. bernicla*. **19.** *B. ruficollis*. **20.** *Alopochen aegyptiaca*. **21.** *Tadorna ferruginea*. **22.** *T. tadorna*. – Scale: 5 cm.



Figs. 23–31. Skulls of Anseriformes, lateral view. – 23. *Aix galericulata*. 24. *Anas penelope*. 25. *A. stepera*. 26. *A. crecca*. 27. *A. platyrhynchos*. 28. *A. acuta*. 29. *A. querquedula*. 30. *A. clypeata*. 31. *Netta rufina*. – Scale: 5 cm.



Figs. 32–39. Skulls of Anseriformes, lateral view. – 32. *Aythya ferina*. 33. *A. nyroca*. 34. *A. fuligula*. 35. *A. marila*. 36. *Somateria mollissima*. 37. *Clangula hyemalis*. 38. *Bucephala clangula* ♂. 39. *B. clangula* ♀. – Scale: 5 cm.



Figs. 40–47. Skulls of Anseriformes, lateral view. – 40. *Melanitta fusca* ♂. 41. *M. fusca* ♀. 42. *M. nigra* ♂. 43. *M. nigra* ♀. 44. *Mergellus albellus*. 45. *Mergus serrator*. 46. *M. merganser*. 47. *Oxyura jamaicensis*. – Scale: 5 cm.

4 References

- BARTHEL, P. H. & HELBIG, A. J. (2005): Artenliste der Vögel Deutschlands. – *Limicola* **19**: 89–111.
- BAUER, H.-G., BEZZEL, E. & FIEDLER, W. (eds.) (2005): Das Kompendium der Vögel Mitteleuropas. Alles über Biologie, Gefährdung und Schutz, vol. 1 Nonpasseriformes – Nichtsperlingsvögel, 808 pp.; Wiesbaden (Aula).
- BROWN, R., FERGUSON, J., LAWRENCE, M. & LEES, D. (2003): Federn, Spuren und Zeichen der Vögel Europas – Ein Feldführer, 3rd edition, 336 pp.; Wiebelsheim (Aula).
- ELLROTT, C. (2008): Vergleichend ökomorphologische Untersuchungen an Schädeln mitteleuropäischer Entenvögel (Anseriformes). – Graduate thesis, Constance University, 67 pp.
- GOODMAN, D. C. & FISHER, H. I. (1962): Functional Anatomy of the feeding apparatus in waterfowl (Aves: Anatidae), 193 pp.; Carbondale (Southern Illinois University Press).
- HOYO, J. DEL, ELLIOTT, A. & SARGATAL, J. (1992) (eds.): Handbook of the birds of the world, vol. 1 (Ostrich to ducks), 696 pp.; Barcelona (Lynx Edicions).
- JANSEN, J. & GESTEL, W. VAN (2009): Skullsite. www.skullsite.com.
- MICKOLEIT, G. (2004): Phylogenetische Systematik der Wirbeltiere, 671 pp.; München (Friedrich Pfeil).

Manuscript received: 9.X.2009, accepted: 26.XI.2009.

Authors' addresses:

CHRISTIAN ELLROTT, Im Brühl 13, 78086 Brigachtal, Germany;
e-mail: christian.ellrott@googlemail.com

Dr. GREGOR SCHMITZ (corresponding author), Botanischer Garten, Universität Konstanz, 78457 Konstanz, Germany;
e-mail: gregor.schmitz@uni-konstanz.de